

PATENT

**APPLICATION FOR UNITED STATES LETTERS PATENT IN THE UNITED
STATES PATENT AND TRADEMARK OFFICE**

(Case No. 98,009-B)

Title: METHOD OF PROMOTING ERYTHROPOIESIS

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METHOD OF PROMOTING ERYTHROPOIESIS

INSA1

5 Cross Reference

INSA2

~~This application is a continuation-in-part of U.S. Provisional Application Serial No. 60/074,106 filed February 9, 1998 and a continuation of U.S. Provisional Application Serial No. 60/111,535 filed December 9, 1998.~~

10 Field of the Invention

The present invention relates to compounds, methods, compositions, and kits for the stimulation of erythropoiesis. More specifically, the present invention relates to methods, compositions, and kits that employ effective amounts of angiotensinogen, angiotensin I (AI), AI analogues, AI fragments and analogues thereof, angiotensin II analogues, AII fragments or analogues thereof or AII AT₂ type 2 receptor agonists for stimulating erythropoiesis.

Background of the Invention

Maintenance of an adequate supply of oxygen to the body tissues is vital to survival. In the United States alone, several million people suffer from anemia secondary to renal failure, chronic inflammatory disease and malignancies (U.S. Patent No. 4,987,121, hereby incorporated by reference in its entirety). Since to a large degree the oxygen-carrying capacity of blood is governed by the concentration of erythrocytes in the blood, the appropriate regulation of erythropoiesis is also crucial.

The early studies of Reissmann (Reissmann, K. R., *Blood* 5:372-80 (1950)) and Erslev (Erslev, A., *Blood* 8:349-57 (1953)) clearly demonstrated the hypoxia-induced stimulation of erythropoietin secretion. When erythropoietin is secreted from the erythropoietin-producing cells in response to hypoxia, it travels through the blood to its target organ, the hematopoietic tissues. In humans, the principal hematopoietic tissue is within the liver before birth, and in the bone marrow after birth. (Id.) There, erythropoietin binds specifically to its receptor on the erythroid progenitor cells called burst forming unit-erythroid (BFU-E) and colony-forming unit-erythroid (CFU-E) and stimulates these cells to proliferate and differentiate (Spivak, J. L., *Int. J. Cell Cloning* 4:139-66 (1986)). BFU-E are the earliest erythroid progenitors and constitute 0.01%, approximately, of the nucleated bone marrow cells. CFU-E are derived from BFU-E, account for about 0.1% of marrow cells, and are much more responsive to erythropoietin than are BFU-E (Spivak, J. L., *supra*); Sawada, K., et al., *J. Clin. Invest.* 80:357-66 (1987)).

The low erythropoietin levels always present appear sufficient for a basal erythropoiesis rate. Relatively small losses of blood do not appear to stimulate increased erythropoietin production (Kickler, T. S., et al., *J. Am. Med. Assoc.* 260:65-7 (1988)). It is only after a major blood loss that there is an increased production of erythropoietin and rate of erythropoiesis.

It has been well-established that the majority of patients with renal insufficiency and anemia have serum erythropoietin levels well below what would be expected for the degree of anemia (Caro, J., et al., *J. Lab. Clin. Med.* 93:449-58 (1979); Radtke, H. W., et al., *Blood* 54:877-84 (1979); Chandra, M., et al., *J. Pediatr.* 113:1015-21 (1988)),

although they can still respond to hypoxia with an increase in serum erythropoietin levels (Radtke, H. W., et al., *Blood* 54:877-84 (1979); Chandra, M., et al., *J Pediatr* 113:1015-21 (1988)). However, this markedly blunted erythropoietin response substantially contributes to the pathogenesis of the anemia (Eschbach, J. W., et al., *Am J Kid Dis* 11:203-9 (1988)). As a result, patients suffering from chronic renal failure and end-stage renal disease, or those undergoing renal transplantation, develop severe anemia and require regular blood transfusions (Royet, U.S. Patent No. 5,482,924).

The use of recombinant human erythropoietin has facilitated treatment of these patients. However, recombinant erythropoietin treatment is extremely costly, and methods that augment the effect of erythropoiesis will permit the use of smaller doses of erythropoietin, and thus will decrease treatment costs. Additionally, increasing the rate of erythropoiesis would significantly improve clinical benefits for the treatment of congenital or acquired aplastic or hypoplastic anemia associated with chronic renal failure, end-stage renal disease, renal transplantation, cancer, AIDS, chemotherapy, radiotherapy, bone marrow transplantation and chronic diseases.

Summary of the Invention

In one aspect, the present invention provides compounds and methods for the augmentation of erythropoiesis by potentiating erythropoietin-induced differentiation with angiotensinogen, angiotensin I (AI), AI analogues, AI fragments and analogues thereof, angiotensin II analogues, AII fragments or analogues thereof or AII AT₂ type 2 receptor agonists as a therapeutic adjunct.

In another aspect, the present invention provides pharmaceutical compositions comprising angiotensinogen, angiotensin I (AI), AI analogues, AI fragments and analogues thereof, angiotensin II analogues, AII fragments or analogues thereof or AII AT₂ type 2 receptor agonists together with erythropoietin and a pharmaceutically acceptable carrier.

In a further aspect, the present invention provides kits for promoting erythropoiesis, wherein the kits comprise an effective amount of angiotensinogen, AI, AI analogues, AI fragments and analogues thereof, AII analogues, AII fragments or analogues thereof or AII AT₂ type 2 receptor agonists, and instructions for using the amount effective of active agent as a therapeutic adjunct to erythropoietin treatment.

In another aspect, the invention provides improved cell culture medium for promoting erythropoiesis, comprising adding an effective amount of the active agents of the invention to promote erythropoiesis.

The methods and kits of the present invention are clinically useful as a therapeutic adjunct for increasing red blood cell production in treating congenital or acquired aplastic or hypoplastic anemia.

Brief Description of the Figures

Figure 1 is a graph showing the effect of 1GD on formation of human burst-forming units-erythroid.

Figure 2 is a graph showing the effect of 24B on formation of human burst-forming units-erythroid.

Figure 3 is a graph showing the effect of 2GD on formation of human burst-forming units-erythroid.

Figure 4 is a graph showing the effect of 5GD on formation of human burst-forming units-erythroid.

5 **Figure 5** is a graph showing the effect of AII(1-7) on formation of human burst-forming units-erythroid.

Figure 6 is a graph showing the effect of AII on formation of human burst-forming units-erythroid.

10 **Detailed Description of the Preferred Embodiments**

All cited patents, patent applications and references are hereby incorporated by reference in their entirety.

As defined herein, the term "erythropoiesis" refers to red blood cell production.

As defined herein, "augmentation of erythropoiesis" may occur either by direct
15 stimulation of erythroid production, by increasing erythropoietin production, or by any other mechanism.

Unless otherwise indicated, the term "active agents" as used herein refers to the group of compounds comprising angiotensinogen, angiotensin I (AI), AI analogues, AI fragments and analogues thereof, angiotensin II analogues, AII fragments or analogues
20 thereof and AII AT₂ type 2 receptor agonists.

The biological formation of angiotensin is initiated by the action of renin on the plasma substrate angiotensinogen (*Circulation Research* 60:786-790 (1987); Clouston et al., *Genomics* 2:240-248 (1988); Kageyama et al., *Biochemistry* 23:3603-3609; Ohkubo

et al., *Proc. Natl. Acad. Sci.* 80:2196-2200 (1983); all references hereby incorporated in their entirety). The substance so formed is a decapeptide called angiotensin I (AI) which is converted to AII by the converting enzyme angiotensinase which removes the C-terminal His-Leu residues from AI (Asp-Arg-Val-Tyr-Ile-His-Pro-Phe-His-Leu [SEQ ID NO:37]). AII is a known pressor agent and is commercially available.

Angiotensin-converting enzyme (ACE) inhibitors have been observed to exacerbate anemia in patients with chronic renal failure and end-stage renal disease (ESRD), as well as in renal transplant recipients (Cruz et al., *Am. J. Kidney Diseases* 28:535-540 (1996); herein incorporated by reference in its entirety). ACE inhibitors appear to induce anemia by decreasing red blood cell production (*Id.*) Some data exist which suggest that ACE inhibitors may reduce red blood cell production by inhibiting angiotensin-mediated erythropoietin synthesis. (Hirikata et al., *Clin. Nephrol.* 26:27-32 (1986); Gould et al., *J. Lab. Clin. Med.* 96:523-534 (1980); Conlon et al., *Transplantation* 56:217-219 (1993)). However, other studies show that ACE inhibitors do not inhibit erythropoietin synthesis, and suggest that angiotensin does not play a role in erythropoiesis. (Cruz et al., 1996; Anderson et al., *Biology of the Neonate* 71:194-197 (1997); Shand et al., *J. Hum. Hypertension* 9:233-235 (1995); Julian et al., *Kidney Int.* 46:1397-1403 (1994); Gaston et al., *Transplant Proc.* 25:1029-1031 (1993); Islam et al., *Transplant Int.* 3:222-225 (1990); Rostaing et al., *Transplant Proc.* 26:280-281 (1994)). Thus, it is unclear whether AII, or any AII analogues and fragments, stimulate production of erythropoietin.

A recent study suggests that activation of the AT1 receptor with angiotensin II enhances erythropoietin-stimulated human erythroid proliferation *in vitro*. (Mrug et al.,

J. Clin. Invest. 100 (9):2310-2314 (1997). Previous studies have indicated that slow infusion of angiotensin II in dogs (Fisher et al., in Annals New York Academy of Sciences, pp. 308-317: 1968) and mice (Fisher et al., *J. Pharmacol. and Exper. Therapeutics* 157:618-625, 1967) led to increased erythropoietin production, while
5 injections of angiotensin II into mice and rats did not lead to such an increase. (Mann et al., P.S.E.B.M. 121:1152-1154 (1966); Bilsel et al., P.S.E.B.M. 114:475-479 (1963)). Thus, there is some evidence that angiotensin II (AII) stimulates erythropoiesis *in vitro*, while the *in vivo* data is unclear. However, there is no data suggesting whether any AII analogues and fragments also stimulate erythropoiesis, whether *in vitro* or *in vivo*. For
10 example, data suggests that the AII fragment AII(1-7) acts through a receptor(s) that is distinct from the AT1 and AT2 receptors which modulate AII activity. (Ferrario et al., J. Am. Soc. Nephrol. 9:1716-1722 (1998); Iyer et al., Hypertension 31:699-705 (1998); Freeman et al., Hypertension 28:104 (1996); Ambuhl et al., Brain Res. Bull. 35:289 (1994). Thus, the stimulatory effect of AII through activation of the AT1 receptor
15 reported by Mrug et al. (see above) does not shed any light on the potential stimulatory effect of AII(1-7).

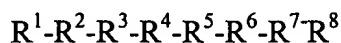
U.S. Patent No. 5,015,629 to DiZerega (the entire disclosure of which is hereby incorporated by reference) describes a method for increasing the rate of healing of wound tissue, comprising the application to such tissue of angiotensin II (AII) in an amount
20 which is sufficient for said increase. The application of AII to wound tissue significantly increases the rate of wound healing, leading to a more rapid re-epithelialization and tissue repair. The term AII refers to an octapeptide present in humans and other species having the sequence Asp-Arg-Val-Tyr-Ile-His-Pro-Phe [SEQ ID NO:1]. The use of AII

analogues and fragments, AT2 agonists, as well as AIII and AIII analogues and fragments in wound healing has also been described. (U.S. Patent No. 5,629,292; U.S. Patent No. 5,716,935; WO 96/39164; all references herein incorporated by reference in their entirety.)

5 A peptide agonist selective for the AT2 receptor (AII has 100 times higher affinity for AT2 than AT1) has been identified. This peptide is p-aminophenylalanine6-AII ["(p-NH₂-Phe)6-AII"], Asp-Arg-Val-Tyr-Ile-Xaa-Pro-Phe [SEQ ID NO.36] wherein Xaa is p-NH₂-Phe (Speth and Kim, BBRC 169:997-1006 (1990)). This peptide gave binding characteristics comparable to AT2 antagonists in the experimental models tested
10 (Catalioto, et al., *Eur. J. Pharmacol.* 256:93-97 (1994); Bryson, et al., *Eur. J. Pharmacol.* 225:119-127 (1992)).

Many studies have focused upon AII(1-7) (AII residues 1-7) or other fragments of AII to evaluate their activity. AII(1-7) elicits some, but not the full range of effects elicited by AII. Pfeilschifter, et al., *Eur. J. Pharmacol.* 225:57-62 (1992); Jaiswal, et al.,
15 *Hypertension* 19 (Supp. II):II-49-II-55 (1992); Edwards and Stack, *J. Pharmacol. Exper. Ther.* 266:506-510 (1993); Jaiswal, et al., *J. Pharmacol. Exper. Ther.* 265:664-673 (1991); Jaiswal, et al., *Hypertension* 17:1115-1120 (1991); Portsi, et al., *Br. J. Pharmacol.* 111:652-654 (1994).

The active AII analogues, fragments of AII and analogues thereof of particular
20 interest in accordance with the present invention are characterized as comprising a sequence consisting of at least three contiguous amino acids of groups R¹-R⁸ in the sequence of general formula I



in which R¹ and R² together form a group of formula



wherein X is H or a one to three peptide group,

R^A is suitably selected from Asp, Glu, Asn, Acpc (1-aminocyclopentane
5 carboxylic acid), Ala, Me²Gly, Pro, Bet, Glu(NH₂), Gly, Asp(NH₂) and Suc,

R^B is suitably selected from Arg, Lys, Ala, Orn, Ser(Ac), Sar, D-Arg and
D-Lys;

R³ is selected from the group consisting of Val, Ala, Leu, Lys, norLeu, Ile,
10 Gly, Pro, Aib, Acpc and Tyr;

R⁴ is selected from the group consisting of Tyr, Tyr(PO₃)₂, Thr, Ser, Ala,
homoSer and azaTyr;

R⁵ is selected from the group consisting of Ile, Ala, Leu, norLeu, Val and
15 Gly;

R⁶ is His, Arg or 6-NH₂-Phe;

15 R⁷ is Pro or Ala; and

R⁸ is selected from the group consisting of Phe, Phe(Br), Ile and Tyr,
excluding sequences including R⁴ as a terminal Tyr group.

Compounds falling within the category of AT2 agonists useful in the practice of
the invention include the AII analogues set forth above subject to the restriction that R⁶ is
20 p-NH₂-Phe.

Particularly preferred combinations for R^A and R^B are Asp-Arg, Asp-Lys, Glu-
Arg and Glu-Lys. Particularly preferred embodiments of this class include the following:
AIII, Arg-Val-Tyr-Ile-His-Pro-Phe [SEQ ID NO:2]; AII(3-8), also known as des1-AIII

or AIV, Val-Tyr-Ile-His-Pro-Phe [SEQ ID NO:3]; AII(1-7), Asp-Arg-Val-Tyr-Ile-His-Pro [SEQ ID NO:4]; AII(2-7), Arg-Val-Tyr-Ile-His-Pro [SEQ ID NO:5]; AII(3-7), Val-Tyr-Ile-His-Pro [SEQ ID NO:6]; AII(5-8), Ile-His-Pro-Phe [SEQ ID NO:7]; AII(1-6), Asp-Arg-Val-Tyr-Ile-His [SEQ ID NO:8]; AII(1-5), Asp-Arg-Val-Tyr-Ile [SEQ ID NO:9]; AII(1-4), Asp-Arg-Val-Tyr [SEQ ID NO:10]; and AII(1-3), Asp-Arg-Val [SEQ ID NO:11]. Other preferred embodiments include: Arg-norLeu-Tyr-Ile-His-Pro-Phe [SEQ ID NO:12] and Arg-Val-Tyr-norLeu-His-Pro-Phe [SEQ ID NO:13]. Still another preferred embodiment encompassed within the scope of the invention is a peptide having the sequence Asp-Arg-Pro-Tyr-Ile-His-Pro-Phe [SEQ ID NO:31]. AII(6-8), His-Pro-Phe [SEQ ID NO:14] and AII(4-8), Tyr-Ile-His-Pro-Phe [SEQ ID NO:15] were also tested and found not to be effective.

A further class of particularly preferred compounds in accordance with the present invention consists of those with the following general structure:

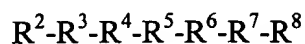
ASP-ARG-R1-R2-R3-R4-PRO-R5

wherein R1 is selected from the group consisting of Val, Pro, and Lys;
R2 is selected from the group consisting of Tyr, Tyr (PO₃)₂ and Ala;
R3 is selected from the group consisting of Ile, Val, Leu, norLeu and Ala;
R4 is selected from the group consisting of His and Arg; and
R5 is either Phe or is absent,
and wherein the active agent is not AII.

Preferred embodiments of this class of the invention include SEQ ID NO:4, SEQ ID NO:19, SEQ ID NO:26, SEQ ID NO:27, SEQ ID NO:31, SEQ ID NO: 32, SEQ ID NO: 34; SEQ ID NO:38, SEQ ID NO:39, and SEQ ID NO. 40. Particularly preferred

embodiments of this class include SEQ ID NO:4, SEQ ID NO. 31, SEQ ID NO 38, and SEQ ID NO. 39.

Another class of compounds of particular interest in accordance with the present invention are those of the general formula II



in which R^2 is selected from the group consisting of H, Arg, Lys, Ala, Orn, Ser(Ac), Sar, D-Arg and D-Lys;

R^3 is selected from the group consisting of Val, Ala, Leu, norLeu, Ile, Gly, Pro, Aib, Acpc and Tyr;

R^4 is selected from the group consisting of Tyr, Tyr(PO₃)₂, Thr, Ser, homoSer and azaTyr;

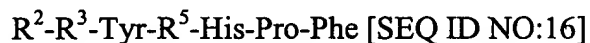
R^5 is selected from the group consisting of Ile, Ala, Leu, norLeu, Val and Gly;

R^6 is His, Arg or 6-NH₂-Phe;

R^7 is Pro or Ala; and

R^8 is selected from the group consisting of Phe, Phe(Br), Ile and Tyr.

A particularly preferred subclass of the compounds of general formula II has the formula



wherein R^2 , R^3 and R^5 are as previously defined. Particularly preferred is angiotensin III of the formula Arg-Val-Tyr-Ile-His-Pro-Phe [SEQ ID NO:2]. Other

preferred compounds include peptides having the structures Arg-Val-Tyr-Gly-His-Pro-Phe [SEQ ID NO:17] and Arg-Val-Tyr-Ala-His-Pro-Phe [SEQ ID NO:18]. The fragment AII(4-8) was ineffective in repeated tests; this is believed to be due to the exposed tyrosine on the N-terminus.

5 In the above formulas, the standard three-letter abbreviations for amino acid residues are employed. In the absence of an indication to the contrary, the L-form of the amino acid is intended. Other residues are abbreviated as follows:

TABLE 1

Abbreviation for Amino Acids

Me ² Gly	N,N-dimethylglycyl
Bet	1-carboxy-N,N,N-trimethylmethanaminium hydroxide inner salt (betaine)
Suc	Succinyl
Phe(Br)	p-bromo-L-phenylalanyl
azaTyr	aza- α' -homo-L-tyrosyl
Acpc	1-aminocyclopentane carboxylic acid
Aib	2-aminoisobutyric acid
Sar	N-methylglycyl (sarcosine)

10 It has been suggested that AII and its analogues adopt either a *gamma* or a *beta* turn (Regoli, et al., Pharmacological Reviews 26:69 (1974)). In general, it is believed that neutral side chains in position R³, R⁵ and R⁷ may be involved in maintaining the appropriate distance between active groups in positions R⁴, R⁶ and R⁸ primarily responsible for binding to receptors and/or intrinsic activity. Hydrophobic side chains in

positions R^3 , R^5 and R^8 may also play an important role in the whole conformation of the peptide and/or contribute to the formation of a hypothetical hydrophobic pocket.

Appropriate side chains on the amino acid in position R^2 may contribute to affinity of the compounds for target receptors and/or play an important role in the conformation of the peptide. For this reason, Arg and Lys are particularly preferred as R^2 .

For purposes of the present invention, it is believed that R^3 may be involved in the formation of linear or nonlinear hydrogen bonds with R^5 (in the gamma turn model) or R^6 (in the beta turn model). R^3 would also participate in the first turn in a beta antiparallel structure (which has also been proposed as a possible structure). In contrast to other positions in general formula I, it appears that beta and gamma branching are equally effective in this position. Moreover, a single hydrogen bond may be sufficient to maintain a relatively stable conformation. Accordingly, R^3 may suitably be selected from Val, Ala, Leu, norLeu, Ile, Gly, Pro, Aib, Acpc and Tyr. Furthermore, Lys has surprisingly been found to be suitable at R^3 (see Examples).

With respect to R^4 , conformational analyses have suggested that the side chain in this position (as well as in R^3 and R^5) contribute to a hydrophobic cluster believed to be essential for occupation and stimulation of receptors. Thus, R^4 is preferably selected from Tyr, Thr, Tyr (PO_3)₂, homoSer, Ser and azaTyr. Furthermore, Ala has surprisingly been found to be suitable at the R^4 position (See Examples). In this position, Tyr is particularly preferred as it may form a hydrogen bond with the receptor site capable of accepting a hydrogen from the phenolic hydroxyl (Regoli, et al. (1974), *supra*).

In position R⁵, an amino acid with a β aliphatic or alicyclic chain is particularly desirable. Therefore, while Gly is suitable in position R⁵, it is preferred that the amino acid in this position be selected from Ile, Ala, Leu, norLeu, Gly and Val.

In the AI and AII analogues, fragments and analogues of fragments of particular interest in accordance with the present invention, R⁶ is His, Arg or 6-NH₂-Phe. The unique properties of the imidazole ring of histidine (e.g., ionization at physiological pH, ability to act as proton donor or acceptor, aromatic character) are believed to contribute to its particular utility as R⁶. For example, conformational models suggest that His may participate in hydrogen bond formation (in the *beta* model) or in the second turn of the antiparallel structure by influencing the orientation of R⁷. Similarly, it is presently considered that R⁷ should be Pro in order to provide the most desirable orientation of R⁸. In position R⁸, both a hydrophobic ring and an anionic carboxyl terminal appear to be particularly useful in binding of the analogues of interest to receptors; therefore, Tyr and especially Phe are preferred for purposes of the present invention.

Analogues of particular interest include the following:

TABLE 2

Angiotensin II Analogues

AII Analogue Name	Amino Acid Sequence	Sequence Identifier
Analogue 1	Asp-Arg-Val-Tyr-Val-His-Pro-Phe	SEQ ID NO: 19
Analogue 2	Asn-Arg-Val-Tyr-Val-His-Pro-Phe	SEQ ID NO: 20
Analogue 3	Ala-Pro-Gly-Asp-Arg-Ile-Tyr-Val-His-Pro-Phe	SEQ ID NO: 21
Analogue 4	Glu-Arg-Val-Tyr-Ile-His-Pro-Phe	SEQ ID NO: 22
Analogue 5	Asp-Lys-Val-Tyr-Ile-His-Pro-Phe	SEQ ID NO: 23
Analogue 6	Asp-Arg-Ala-Tyr-Ile-His-Pro-Phe	SEQ ID NO: 24
Analogue 7	Asp-Arg-Val-Thr-Ile-His-Pro-Phe	SEQ ID NO: 25
Analogue 8	Asp-Arg-Val-Tyr-Leu-His-Pro-Phe	SEQ ID NO: 26
Analogue 9	Asp-Arg-Val-Tyr-Ile-Arg-Pro-Phe	SEQ ID NO: 27

Analogue 10	Asp-Arg-Val-Tyr-Ile-His-Ala-Phe	SEQ ID NO: 28
Analogue 11	Asp-Arg-Val-Tyr-Ile-His-Pro-Tyr	SEQ ID NO: 29
Analogue 12	Pro-Arg-Val-Tyr-Ile-His-Pro-Phe	SEQ ID NO: 30
Analogue 13	Asp-Arg-Pro-Tyr-Ile-His-Pro-Phe	SEQ ID NO: 31
Analogue 14	Asp-Arg-Val-Tyr(Po ₃) ₂ -Ile-His-Pro-Phe	SEQ ID NO: 32
Analogue 15	Asp-Arg-norLeu-Tyr-Ile-His-Pro-Phe	SEQ ID NO: 33
Analogue 16	Asp-Arg-Val-Tyr-norLeu-His-Pro-Phe	SEQ ID NO: 34
Analogue 17	Asp-Arg-Val-homoSer-Tyr-Ile-His-Pro-Phe	SEQ ID NO: 35

The polypeptides of the instant invention may be synthesized by a wide variety of methods such as those set forth in J. M. Stewart and J. D. Young, Solid Phase Peptide Synthesis, 2nd ed., Pierce Chemical Co., Rockford, Ill. (1984) and J. Meienhofer, Hormonal Proteins and Peptides, Vol. 2, Academic Press, New York, (1973) for solid phase synthesis and E. Schroder and K. Lubke, The Peptides, Vol. 1, Academic Press, New York, (1965) for solution synthesis. The disclosures of the foregoing treatises are incorporated by reference herein.

In general, these methods involve the sequential addition of protected amino acids to a growing peptide chain (U.S. Patent No. 5,693,616, herein incorporated by reference in its entirety). Normally, either the amino or carboxyl group of the first amino acid and any reactive side chain group are protected. This protected amino acid is then either attached to an inert solid support, or utilized in solution, and the next amino acid in the sequence, also suitably protected, is added under conditions amenable to formation of the amide linkage. After all the desired amino acids have been linked in the proper sequence, protecting groups and any solid support are removed to afford the crude polypeptide. The polypeptide is desalted and purified, preferably chromatographically, to yield the final product.

In one aspect, the present invention provides methods for the augmentation of erythropoiesis by potentiating erythropoietin-induced differentiation with

angiotensinogen, angiotensin I (AI), AI analogues, AI fragments and analogues thereof, angiotensin II analogues, AII fragments or analogues thereof or AII AT₂ type 2 receptor agonists (ie: "active agents") as a therapeutic adjunct to erythropoietin treatment. The methods and kits of the present invention are clinically useful as a therapeutic adjunct for
5 increasing red blood cell production in treating congenital or acquired aplastic or hypoplastic anemia.

The use of erythropoietin to promote erythropoiesis is well known in the art, as exemplified by Royet et al., U.S. Patent No. 5,482,924; Goldberg et al., U.S. Patent No. 5,188,828; Vance et al., U.S. Patent No. 5,541,158; and Baertschi et al., U.S. Patent No.
10 4,987,121, all references hereby incorporated in their entirety. The erythropoietin dosage regimen may vary widely, but can be determined routinely by a physician using standard methods. Dosage levels of the order of between about 1 EPO unit/kg and about 5,000 EPO units/kg body weight are useful for all methods of use disclosed herein.

In one embodiment, the effects of the active agents on the growth of erythroid progenitors *in vitro* are tested using the colony formation assay. The assay consists of
15 growing erythroid progenitor cells in a semi-solid medium (methylcellulose) for two weeks (Yu et al., U.S. Patent No. 5,032,507). Conditioned medium consisting of phytohemagglutinin-treated lymphocytes (PHA-LCM) is supplemented with erythropoietin and preferably, between about 0.1 ng/ml and about 10 mg/ml of the active
20 agents.

The growth of erythroid precursors termed BFU-E (burst forming units erythroid) is monitored by identification and counting of the colonies under an inverted microscope as well as by staining of colonies (Yu et al., U.S. Patent No. 5,032,507). The number of

mixed colonies represents the number of earlier progenitor cells (containing erythroid as well as one or more other lineage cells).

In another embodiment, erythropoiesis is augmented *ex vivo* by obtaining a sample of bone marrow cells, as is known in the art, potentiating erythropoietin-induced differentiation with the active agents of the invention and infusing the treated cells back into the patient.

In a preferred embodiment, bone marrow cells are isolated from peripheral blood samples via standard techniques (U.S. Patent Nos. 4,987,121, 5,104,653; hereby incorporated by reference in their entirety). 2×10^6 bone marrow cells are seeded in culture dishes in appropriate medium, such as modified Dulbecco's medium (IMDM) supplemented with (final concentration): horse serum (15%), fetal calf serum (5%), Fe saturated transferrin (0.4 mg/ml) hydrocortisone, penicillin 100 u/ml, and streptomycin (0.1 mg/ml) (Royet et al., U.S. Patent No. 5,482,924). An adherent cell monolayer is formed. After 15 days, the non adherent cells are removed and fresh bone marrow is reseeded in the presence of 0.1 U/ml of erythropoietin (EPO) and, preferably, between about 0.1 ng/ml and about 10 mg/ml of the active agents of the invention. The cells are expanded for a period of between 2 and 21 days with subsequent medium changes as required. Prior to reinfusion into the subject the cells are examined microscopically to verify the absence of contamination. The cells are rinsed to remove all traces of culture fluid, resuspended in an appropriate medium and then pelleted and rinsed several times. After the final rinse, the cells are resuspended at between 0.7×10^6 and 50×10^6 cells per ml in an appropriate medium and reinfused into a subject. Erythropoiesis is monitored by

red cell count or hemoglobin concentration with time (Yu et al., U.S. Patent No. 5,032,507, herein incorporated by reference in its entirety).

For use in increasing erythropoiesis *in vivo*, the active agents may be administered by any suitable route, including orally, parentally, by inhalation spray, rectally, 5 transdermally, or topically in dosage unit formulations containing conventional pharmaceutically acceptable carriers, adjuvants, and vehicles. The term parenteral as used herein includes, subcutaneous, intravenous, intra-arterial, intramuscular, intrasternal, intratendinous, intraspinal, intracranial, intrathoracic, infusion techniques or intraperitoneally.

10 Transdermal means including, but not limited to, transdermal patches may be utilized to deliver the active agents to the treatment site. Transdermal formulations may be prepared by incorporating the active agent in a thixotropic or gelatinous carrier including, but not limited to a cellulose medium, e.g., methyl cellulose or hydroxyethyl cellulose, with the resulting formulation then being packed in a transdermal device 15 adapted to be secured in dermal contact with the skin of a wearer.

The active agents may be made up in a solid form (including granules, powders or suppositories) or in a liquid form (e.g., solutions, suspensions, or emulsions). The active agents may be subjected to conventional pharmaceutical operations such as sterilization and/or may contain conventional adjuvants, such as preservatives, stabilizers, wetting 20 agents, emulsifiers, buffers etc.

While the active agents can be administered as the sole active pharmaceutical agent, they can also be used in combination with one or more other compounds. When administered as a combination, the active agent(s) and compound(s) can be formulated as

separate compositions that are given at the same time or different times, or the active agent(s) and compound(s) can be given as a single composition.

For administration, the active agents are ordinarily combined with one or more adjuvants appropriate for the indicated route of administration. The active agents may be admixed with lactose, sucrose, starch powder, cellulose esters of alkanolic acids, stearic acid, talc, magnesium stearate, magnesium oxide, sodium and calcium salts of phosphoric and sulphuric acids, acacia, gelatin, sodium alginate, polyvinylpyrrolidone, and/or polyvinyl alcohol, and tableted or encapsulated for conventional administration. Alternatively, the compounds of this invention may be dissolved in saline, water, polyethylene glycol, propylene glycol, carboxymethyl cellulose colloidal solutions, ethanol, corn oil, peanut oil, cottonseed oil, sesame oil, tragacanth gum, and/or various buffers. Other adjuvants and modes of administration are well known in the pharmaceutical art. The carrier or diluent may include time delay material, such as glyceryl monostearate or glyceryl distearate alone or with a wax, or other materials well known in the art.

Formulations suitable for topical administration include liquid or semi-liquid preparations suitable for penetration through the skin (*e.g.*, liniments, lotions, ointments, creams, or pastes) and drops suitable for administration to the eye, ear, or nose.

The dosage regimen for augmenting erythropoiesis with the active agents is based on a variety of factors, including the type of injury, the age, weight, sex, medical condition of the individual, the severity of the condition, the route of administration, and the particular compound employed. Thus, the dosage regimen may vary widely, but can be determined routinely by a physician using standard methods. Dosage levels of the

order of between 0.1 ng/kg and 10 mg/kg of the active agents per body weight are useful for all methods of use disclosed herein.

The treatment regime will vary depending on the disease being treated, based on a variety of factors, including the type of injury, the age, weight, sex, medical condition of the individual, the severity of the condition, the route of administration, and the particular compound employed.

In a preferred embodiment of the present invention, the active agents are administered intravenously. A suitable dose of the active agents is preferably between about 0.1 ng/kg and about 10 mg/kg administered twice daily. For topical administration, the active ingredient may comprise from 0.001% to 10% w/w, *e.g.*, from 1% to 2% by weight of the formulation, although it may comprise as much as 10% w/w, but preferably not more than 5% w/w, and more preferably from 0.1% to 1% of the formulation.

In another aspect, the present invention provides a novel peptide with erythropoiesis-promoting activity, consisting of the peptide with the sequence Asp-Arg-Lys-Tyr-Ile-His-Pro-Phe (SEQ ID NO:39).

A further object of the present invention is to provide pharmaceutical compositions comprising the active agents as an ingredient for use in the method of the invention. The compositions comprise the active agents together with erythropoietin and a pharmaceutically acceptable carrier, this term including any carrier which does not interfere with the effectiveness of the biological activity of the active agents and erythropoietin, and which is not toxic to the host to which it is administered. Dosage and administration of the pharmaceutical compositions will vary depending on the disease being treated, based on a variety of factors, including the type of injury, the age, weight,

sex, medical condition of the individual, the severity of the condition, the route of administration, and the particular compound employed, as above. Thus, the dosage regimen may vary widely, but can be determined routinely by a physician using standard methods.

5 In a further aspect, the present invention provides kits promoting erythropoiesis, wherein the kits comprise an effective amount of the active agent, and instructions for using the amount effective of active agent as a therapeutic adjunct. In a preferred embodiment, the kit further comprises a pharmaceutically acceptable carrier, such as those adjuvants described above. In another preferred embodiment, the kit further
10 comprises a means for delivery of the active agent to a mammal. Such devices include, but are not limited to matrical or micellar solutions, polyethylene glycol polymers, carboxymethyl cellulose preparations, crystalloid preparations (e.g., saline, Ringer's lactate solution, phosphate-buffered saline, etc.), viscoelastics, polyethylene glycols, and polypropylene glycols.

15 In a further preferred embodiment, the kits also comprise an amount of erythropoietin effective to accelerate erythropoiesis.

 In another aspect of the present invention, an improved cell culture medium is provided for the promotion of erythropoiesis, wherein the improvement comprises addition to the cell culture medium of an effective amount of the active agents of the
20 invention. Any cell culture media that can support erythropoiesis can be used with the present invention. Such cell culture media include, but are not limited to Basal Media Eagle, Dulbecco's Modified Eagle Medium, Iscove's Modified Dulbecco's Medium,

McCoy's Medium, Minimum Essential Medium, F-10 Nutrient Mixtures, Opti-MEM® Reduced-Serum Medium, and RPMI Medium, or combinations thereof.

5 The improved cell culture medium can be supplied in either a concentrated (ie: 10X) or non-concentrated form, and may be supplied as a liquid, a powder, or a lyophilizate. The cell culture may be either chemically defined, or may contain a serum supplement. Culture media and serum supplements are commercially available from many sources, such as GIBCO BRL (Gaithersburg, MD) and Sigma (St. Louis, MO).

10 The present invention, by providing methods and pharmaceutical compositions for augmenting erythropoiesis, will greatly increase the clinical benefits of treatment for congenital or acquired aplastic or hypoplastic anemia; amelioration of anemia associated with cancer, AIDS, chemotherapy, radiotherapy, bone marrow transplantation and chronic diseases.

15 The present invention may be better understood with reference to the accompanying examples that are intended for purposes of illustration only and should not be construed to limit the scope of the invention, as defined by the claims appended hereto.

Example 1. AII and AII analogue and fragment effect on erythroid progenitor formation

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CD34+ cells were isolated from human cord blood by immunomagnetic chromatography using an antibody cocktail (Stem Cell Technologies, Vancouver BC) consisting of the following cell surface proteins: glycophorin A, CD56, CD66b, CD3, CD24, CD14, CD2, CD19, and CD16. Enriched cells were cultured at 37° at 5% CO₂ and air for 6 days in medium containing serum-free StemSpan (Stem Cell Technologies),

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3 IU/ml human erythropoietin (EPO), 20 ng/ml stem cell factor, 20 ng/ml interleukin 3, and 20 ng/ml GM-CSF. The cells were harvested, counted and cultured in 96 well plates at a density of 50,000 cells per well in the same medium containing from 0 to 10 µg/ml of AII, AII analogues, or AII fragments. The peptides tested, and the figure showing the data received in those tests, are listed in Table 3. After 3 additional days in suspension culture (day 9), the cells were washed to remove the peptides and cultured to assess colony formation. The culture medium contained 0.9% methylcellulose in Iscove's MDM with 30% fetal calf serum, 1% bovine serum albumin, 10 µM 2-mercaptoethanol, 2mM L-glutamine, 10% Agar Leukocyte Conditioned Medium with 3 IU/ml EPO. On days 2, 4, 9, and 14 the number and size of colonies was assessed as well as the number of BFU-E formed (burst forming units-erythroid) as a measure of erythropoiesis.

Table 3. Designation for Peptides Tested

Name	Abbreviation	Sequence	Figure	SEQ ID
1GD	Ala ⁴ -AII(1-7)	DRVAIHP	1	SEQ ID NO. 18
GSD 24B	Pro ³ -AII	DRPYIHPF	2	SEQ ID NO. 31
2GD	Pro ³ -AII(1-7)	DRPYIHP	3	SEQ ID NO. 38
5GD	Lys ³ -AII(1-7)	DRKYIHP	4	SEQ ID NO 39
AII(1-7)		DRVYIHP	5	SEQ ID NO. 4
AII		DRVYIHPF	6	SEQ ID NO. 1

The data from these experiments are presented in Figures 1-6 and show that each of the peptides tested increased the number of BFU-E formed relative to control where no peptide was added. Each of the peptides also increased the size of the colonies assessed (data not shown). Therefore, these data demonstrate that each of the peptides tested can be used to accelerate erythroid progenitor formation, and thus to promote erythropoiesis.

The methods and kits of the present invention are clinically useful as a therapeutic adjunct for increasing red blood cell production in treating congenital or acquired aplastic or hypoplastic anemia.

The present invention is not limited by the aforementioned particular preferred
5 embodiments. It will occur to those ordinarily skilled in the art that various modifications may be made to the disclosed preferred embodiments without diverting from the concept of the invention. All such modifications are intended to be within the scope of the present invention.